

IN THE DRAWINGS**Amendments To The Drawings:**

The drawings are amended to correct informalities. Particularly, a Replacement Sheet of Figure 1 is submitted herewith so as to be consistent with formal corrections made to the Specification. No new matter is added.

REMARKS

Favorable reconsideration of this application is respectfully requested in view of the above amendments and the following remarks. The specification and drawings have been amended to correct informalities only. Particularly, the specification is amended to revise concentration references from "mg/dL" to "mg/mL" and "ng/mL" respectively, the values of which have been converted accordingly. Further, the Figure 1 is amended to revise the concentration references from "mg/dL" to "mg/mL," where the values reflected in the graph have been converted accordingly. Applicants submit herewith a Substitute Specification and Replacement Drawing of Figure 1 showing these revisions. Claims 1, 8-11, 14 and 16 are amended and are supported for example at page 8, line 3, page 9, line 27, and Examples. Claims 2-7, 13, and 15 are canceled without prejudice or disclaimer. No new matter has been added. Claims 1, 8-12, 14, and 16 are pending.

The issue of priority is raised at page 2 of the Office Action and is addressed as follows. Applicants respectfully submit that foreign priority is not being claimed in this application. (See for example Applicants' Application Data Sheet page no. 4.)

The specification is objected to for informalities. Applicants respectfully submit that the specification has been amended to change concentration reference of "mg/dL" to "mg/mL" and "ng/mL" as suggested. Further, the represented values have been converted to be consistent with the revised concentration designations. Applicants respectfully submit that the specification is in proper form.

Withdrawal of the objection is respectfully requested.

Claims 1-16 are objected to for informalities. Claim 1 has been amended to recite "An immunoassay method," so as to include proper preamble language. Claim 16 has been amended to remove the active step of "combining" to better present a claim directed to a kit. Applicants respectfully submit that the claims are in proper form.

Withdrawal of the objection is respectfully requested.

Claims 13 and 14 are rejected under 35 U.S.C. 112, second paragraph, as being indefinite. Claim 13 is canceled. Claim 14 has been revised to depend upon claim 12. Thus, Applicants respectfully submit that the rejection is no longer applicable.

Withdrawal of the rejection is respectfully requested.

Claims 1-4, 7, and 9-15 are rejected under 35 U.S.C. 103(a) as being unpatentable over Eda et al. (U.S. Patent No. 6,248,597) in combination with Shigenobu et al. (WO 02/018953). Applicants respectfully traverse this rejection to the extent it is maintained.

Claim 1 is directed to an immunoassay method of a prostate-specific antigen (PSA) comprising performing an antigen-antibody reaction in the presence of a copolymer obtained by polymerizing a monomer represented by the following general formula [2] and an aralkyl methacrylate or derivatives thereof. Claim 11 is directed to a kit of reagent for immunoassay of a PSA comprising a reagent containing the same copolymer and a reagent containing an antibody to a prostate-specific antigen or a prostate-specific antigen.

The combination of Eda et al. and Shigenobu et al. do not teach or suggest the features of claims 1 and 11. Eda et al teaches an agglutination immunoassay of an antigenic analyte that includes performing an antigen-antibody reaction in the presence of a microparticle (e.g. latex) and that the antigenic analyte may be a tumor marker such as PSA. (Col. 4, line 67 to Col. 5, line 5; Col. 7, lines 12-19; and Col. 5, line 47.) Shigenobu et al teaches an agglutination immunoassay that allows an antigenic substance in a sample to bind to insoluble carrier particles and allowing an antibody or an antibody complex which reacts specifically to the antigenic substance to bind to the antigenic substance in the presence of a polymer. (Page 2, line 43-50.) The polymer in Shigenobu et al includes either a polymer formed from 2-methacryloyloxyethyl phosphorylcholine as a monomer, or a copolymer obtained by polymerizing the monomer 2-methacryloyloxyethyl phosphorylcholine with another monomer selected from the group consisting of (meth)acrylates such as acrylate ester, a methacrylate ester, butyl methacrylate or styrene derivatives. (Page 5, lines 4-29.)

As recognized in the Office Action, however, the cited references do not teach or suggest the polymer obtained by polymerizing 2-methacryloyloxyethyl phosphorylcholine, namely the monomer of the general formula [2], and benzyl methacrylate. Likewise, the cited references do not teach or suggest the copolymer obtained by polymerizing the monomer of the general formula [2] and an aralkyl methacrylate or derivatives thereof. Furthermore, the cited references do not suggest any motivation to combine their teachings to arrive at the claimed invention. Thus, Eda et al.

and Shigenobu et al. do not teach or suggest the features required by claims 1 and 11. Applicants respectfully submit that claims 1 and 11 and the respective dependent claims are not obvious.

Favorable consideration and withdrawal of the rejection are respectfully requested.

Claims 5 and 6 are rejected under 35 U.S.C. 103(a) as being unpatentable over Eda et al. (U.S. Patent No. 6,248,597) in combination with Shigenobu et al. (WO 02/018953) and further in view of Sakaki et al. (JP 2000-093169, Abstract).

Claims 5 and 6 have been canceled, rendering this rejection moot. Applicants, however, do not concede the correctness of the rejection.

Withdrawal of the rejection is respectfully requested.

In view of the above amendments and remarks, favorable reconsideration is respectfully requested and a Notice of Allowance is respectfully solicited. If any further questions arise regarding this communication, the Examiner is invited to contact the undersigned attorney.



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Respectfully submitted,

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A handwritten signature in dark ink, appearing to read 'Curtis B. Hamre'. The signature is written over a horizontal line.

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DESCRIPTION

AGGLUTINATION ACCELERATOR FOR IMMUNOLOGICAL MEASUREMENT

5 DETAILED DESCRIPTION OF THE INVENTION

PRIOR ART

A method for confirming presence of a target substance in a sample or measuring a concentration of a target substance in a sample by observing formation of agglutination after mixing immobilized antigen or antibody on a proper carrier and a biological origin sample such as serum, plasma, urine, and the like, or a method for confirming presence of a target substance in a sample or measuring a concentration of a target substance in a sample on the basis of turbidity generated by an antigen-antibody reaction, is an assay method known as so-called immunoassay.

In a case of measuring agglutination or turbidity caused by the antigen-antibody reaction, various compounds so-called agglutination accelerator are generally used. The agglutination accelerator has an action to generate more easily an agglutination based on the antigen-antibody reaction. Examples of such agglutination accelerators are, for example, polyethylene glycol, dextran, carboxymethyl cellulose, hydroxyethyl cellulose, and the like, and among them polyethylene glycol is commonly used.

Since polyethyleneglycol is salted out in a solution of high salt concentration, one of major problems is to worsen the accuracy of measurement caused by hightning blank value of the reagent when such solution is used as reagent for

immunoassay.

On the other hand, there is recently a trend that a high sensitive assay method is performed by utilizing agglutination or turbidity generated by the antigen-antibody reaction. As a result, there is a demand for development of the agglutination accelerator, which has an agglutination accelerating effect equal to or stronger than the conventionally used polyethylene glycol; hardly generates non-specific turbidity; and hardly generates salting out even in a solution with a high salt concentration.

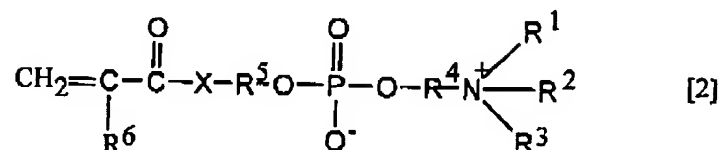
MEANS FOR SOLVING THE PROBLEM

The present invention has been completed considering these situations. An object of the present invention is to provide an immunoassay of prostate-specific antigen (PSA) using an agglutination accelerator, which has an agglutination accelerating effect equal to or stronger than the conventionally known agglutination accelerator; hardly generates non-specific turbidity; and hardly generates salting out even in a solution with a high salt concentration, and a kit of reagent for the immunoassay comprising the reagent containing said agglutination accelerator. Another object of the present invention is to provide the following as the means thereof.

That is, the present invention relates to:

(1) An immunoassay of a prostate-specific antigen comprising performing an antigen-antibody reaction in the presence of a polymer having a monomer unit derived from a monomer represented by the following general formula [2]:

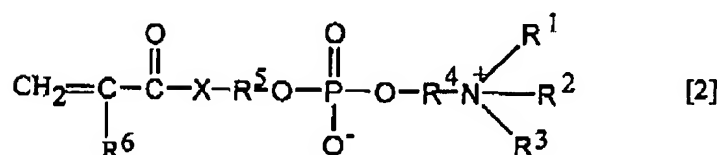
(wherein, R^1 to R^3 are each independently a hydrogen atom



or an alkyl group optionally having a hydroxyl group; R^4 is an alkylene group; R^5 is an alkylene group optionally having a substituent and optionally having an oxygen atom
 5 in the chain; R^6 is a hydrogen atom or a methyl group; and X is an oxygen atom or a -NH- group).

(2) A kit of reagent for immunoassay of a prostate-specific antigen comprising combining a reagent containing:

a copolymer obtained by polymerizing a monomer
 10 represented by the following general formula [2]:



(wherein, R^1 to R^3 are each independently a hydrogen atom or an alkyl group optionally having a hydroxyl group; R^4 is an alkylene group; R^5 is an alkylene group optionally
 15 having a substituent and optionally having an oxygen atom in the chain; R^6 is a hydrogen atom or a methyl group; and X is an oxygen atom or a -NH- group), and a monomer selected from a group consisting of acrylic acid or acrylate ester, methacrylic acid or methacrylate ester, acrylamide or
 20 N-substituted derivatives thereof, methacrylamide or N-substituted derivatives thereof, and styrene or derivatives thereof; and

a reagent containing an antibody to the prostate-specific antigen or a prostate-specific antigen.

Namely, the present inventors extensively studied to find out an immunoassay of PSA using an agglutination
5 accelerator, which has an agglutination accelerating effect equal to or stronger than that of the known agglutination accelerator; hardly generates non-specific turbidity; and hardly generates salting out even in a solution with a high salt concentration, and a kit of reagent for the immunoassay
10 comprising a reagent containing the agglutination accelerator for the immunoassay, and finally found out that the polymer represented by the general formula [2] (hereinafter sometimes abbreviated as the polymer of the present invention) had the intended properties, and thus
15 the present invention was completed.

BRIEF DESCRIPTION OF THE DRAWINGS

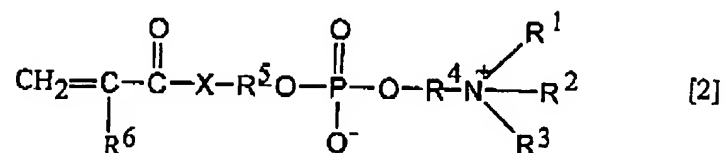
Fig. 1 is a drawing which shows a correlation between
20 CRP concentration (mg/[dL]mL) obtained in Referential Example 5 and CRP concentration (mg/[dL]mL) obtained in Comparative Example 4.

BEST MODE FOR CARRYING OUT THE INVENTION

The polymer used as an agglutination accelerator
25 in the present invention is not particularly limited and may be a homopolymer or a copolymer, usually having a molecular weight of 10,000 to 1,000,000, preferably 10,000 to 500,000, and more preferably 50,000 to 500,000.

More specifically, the polymer preferably includes

a polymer having a monomer unit derived from a monomer represented by the following general formula [2]:



wherein, R¹ to R³ are each independently a hydrogen atom or an alkyl group optionally having a hydroxyl group; R⁴ is an alkylene group; R⁵ is an alkylene group optionally having a substituent and optionally having an oxygen atom in the chain; R⁶ is a hydrogen atom or a methyl group; and X is an oxygen atom or a -NH- group.

In the above-described general formula [2], an alkyl group optionally having a hydroxyl group represented by R¹ to R³ may be linear, branched or cyclic, and usually includes a group having 1-22 carbon atoms, preferably 1 to 20 carbon atoms, more preferably 1 to 2 carbon atoms, and further more preferably 1, and specifically includes such a group as methyl, ethyl, n-propyl, isopropyl, n-butyl, isobutyl, sec-butyl, tert-butyl, n-pentyl, isopentyl, sec-pentyl, tert-pentyl, n-hexyl, isohexyl, sec-hexyl, tert-hexyl, cyclopropyl, cyclohexyl, cyclopentyl, lauryl, myristyl, palmityl and stearyl group, and methyl group, ethyl group and the like are preferable, and methyl group and the like is more preferable.

The alkyl group having a hydroxyl group includes a group in which 1 to 2 hydrogen atoms, preferably one hydrogen atom in the above-described alkyl group are substituted with

a hydroxyl group, and specifically includes a group such as hydroxymethyl, hydroxyethyl, hydroxy-n-propyl, hydroxyisopropyl, hydroxy-n-butyl, hydroxyisobutyl, hydroxy-sec-butyl, hydroxy-tert-butyl, hydroxy-n-pentyl, hydroxyisopentyl, hydroxy-sec-pentyl, hydroxy-tert-pentyl, hydroxy-n-hexyl, hydroxyisohexyl, hydroxy-sec-hexyl, hydroxy-tert-hexyl, hydroxycyclopropyl, hydroxycyclohexyl and hydroxycyclopentyl group, and hydroxymethyl group and hydroxyethyl group are preferable.

10 The alkylene group represented by R^4 includes, for example, an alkylene group having 1 to 6 carbon atoms, preferably 2 to 3 carbon atoms, and may be linear, branched or cyclic, and specifically includes a group such as methylene, ethylene, propylene, trimethylene, butylene, 15 1-ethylethylene, 2-methyltrimethylene, 2-ethyltrimethylene, hexylene, cyclopropylene, cyclobutylene, cyclopentylene and cyclohexylene group, and ethylene group, propylene group and trimethylene group are preferable.

20 In the alkylene group represented by R^5 in the above-described general formula [2], an alkylene group, optionally having a substituent and optionally having an oxygen atom in the chain, which does not have an oxygen atom includes, for example, the group having 1 to 10 carbon atoms, 25 preferably 1 to 6 carbon atoms and more preferably 2 to 6 carbon atoms, which may be linear, branched or cyclic. The alkylene group specifically includes, for example, a group such as methylene, ethylene, propylene, trimethylene, butylene, 1-ethylethylene, 2-methyltrimethylene,

2-ethyltrimethylene, hexylene, cyclopropylene, cyclobutylene, cyclopentylene and cyclohexylene group. The above substituent includes, for example, an alkoxy group having 1 to 6 carbon atoms, preferably 1 to 3 carbon atoms, and may be linear, branched or cyclic, and specifically includes, for example, a group such as methoxy, ethoxy, n-propoxy, isopropoxy, n-butoxy, isobutoxy, sec-butoxy, tert-butoxy, n-pentyloxy, isopentyloxy, sec-pentyloxy, tert-pentyloxy, n-hexyloxy, isohexyloxy, sec-hexyloxy, tert-hexyloxy, cyclopropoxy, cyclohexyloxy and cyclopentyloxy group; and a halogen atom, more specifically fluorine, chlorine, bromine and iodine atom. A group such as ethylene, propylene, trimethylene and butylene group is preferable. When the group has an oxygen atom in the chain, number of said oxygen atom is 1 to 5, preferably 1 to 3, and the group specifically includes, for example, a group represented by $-(C_2H_4O)_n-C_2H_4-$ (wherein, n is an integer of 1 to 5).

When the polymer, having a monomer unit derived from a monomer represented by the above-described general formula [2], is a copolymer, a monomer unit other than the monomer unit derived from a monomer represented by the above-described general formula [2] includes an unit derived from a monomer selected from, for example, acrylic acid or acrylate ester, methacrylic acid or methacrylate ester, acrylamide or N-substituted derivatives thereof, methacrylamide or N-substituted derivatives thereof, and styrene or derivatives thereof. Besides, two or more kinds of these monomer units may be contained in a copolymer.

Said acrylate ester includes alkyl acrylate, aralkyl acrylate and the like; the methacrylate ester include alkyl methacrylate ester, aralkyl methacrylate ester and the like; the N-substituted acrylamides include N-alkyl
5 acrylamide and N-aralkyl acrylamide; the N-substituted methacrylamides include N-alkyl methacrylamide and N-aralkyl methacrylamide; and the styrene derivatives include α -methylstyrene, substituted styrene, substituted α -methylstyrene and the like.

10 The alkyl group in the above-described alkyl acrylate ester, alkyl methacrylate ester, N-alkyl acrylamides and N-alkyl methacrylamides may be linear, branched or cyclic, and usually has 1-6 carbon atoms, more preferably 1-4 carbon atoms, and specifically includes, for
15 example, a group such as methyl, ethyl, n-propyl, isopropyl, n-butyl, isobutyl, sec-butyl, tert-butyl, n-pentyl, isopentyl, sec-pentyl, tert-pentyl, n-hexyl, isohexyl, sec-hexyl, tert-hexyl, cyclopropyl, cyclohexyl and cyclopentyl group. Said alkyl group may have a substituent,
20 which includes, for example, trialkylammonio group (wherein, an alkyl group includes, for example, methyl group, ethyl group, n-propyl group and isopropyl group having 1-3 carbon atoms. When an alkyl group has a trialkylammonio substituent, said substituent is positively charged and thus usually bound
25 with a counter ion. Such a counter ion includes a halogen ion such as fluorine, chlorine, bromine and iodine ion).

The aralkyl group in aralkyl acrylate ester, aralkyl methacrylate ester, N-aralkyl acrylamides and N-aralkyl methacrylamides includes an aralkyl group having 7 to 10

carbon atoms, and specifically includes, for example, a group such as benzyl, phenylethyl, phenylpropyl and phenylbutyl groups.

The substituent which may be possessed by styrene
5 or α -methylstyrene includes, for example, a linear, branched
or cyclic alkyl group usually having 1 to 6 carbon atoms,
more preferably 1 to 4 carbon atoms (specifically including,
for example, a group such as methyl, ethyl, n-propyl,
isopropyl, n-butyl, isobutyl, sec-butyl, tert-butyl,
10 n-pentyl, isopentyl, sec-pentyl, tert-pentyl, n-hexyl,
isohexyl, sec-hexyl, tert-hexyl, cyclopropyl, cyclohexyl
and cyclopentyl group); a linear, branched or cyclic alkoxy
group usually having 1 to 6 carbon atoms, more preferably
1 to 4 carbon atoms (specifically including, for example,
15 a group such as methoxy, ethoxy, n-propoxy, isopropoxy,
n-butoxy, isobutoxy, sec-butoxy, tert-butoxy, n-pentyloxy,
isopentyloxy, sec-pentyloxy, tert-pentyloxy, n-hexyloxy,
isohexyloxy, sec-hexyloxy, tert-hexyloxy, cyclopropoxy,
cyclohexyloxy and cyclopentyloxy group); a halogen atom such
20 as fluorine, chlorine, bromine and iodine atom; a carboxyl
group; a hydroxyl group and an amino group.

Specific examples of these monomers include, for
example, methacrylic acid, methyl methacrylate, ethyl
methacrylate, propyl methacrylate, butyl methacrylate,
25 2-ethylhexyl methacrylate, lauryl methacrylate, stearyl
methacrylate, 2-trimethylammonioethyl methacrylate,
benzyl methacrylate, phenylethyl methacrylate, acrylic acid,
methyl acrylate, ethyl acrylate, butyl acrylate,
2-ethylhexyl acrylate, lauryl acrylate, stearyl acrylate,

2-trimethylammonioethyl acrylate, benzyl acrylate,
 phenylethyl acrylate; acrylamide, N-methylacrylamide,
 N-ethylacrylamide, N-butylacrylamide,
 N-2-ethylhexylacrylamide, N-laurylacrylamide,
 5 N-stearylacrylamide, N-2-trimethylammonioethylacrylamide,
 N-benzylacrylamide, N-phenylethylacrylamide;
 methacrylamide, N-methylmethacrylamide,
 N-ethylmethacrylamide, N-butylmethacrylamide,
 N-2-ethylhexylmethacrylamide, N-laurylmethacrylamide,
 10 N-stearylmethacrylamide,
 N-2-trimethylammonioethylmethacrylamide,
 N-benzylmethacrylamide, N-phenylethylmethacrylamide;
 styrene, carboxystyrene, hydroxystyrene, aminostyrene,
 methylstyrene, ethylstyrene, methoxystyrene,
 15 ethoxystyrene, chlorostyrene, bromostyrene, α
 -methylstyrene, α -methyl-carboxystyrene, α
 -methyl-hydroxystyrene, α -methyl-aminostyrene, α
 -methyl-methylstyrene, α -methyl-ethylstyrene, α
 -methyl-methoxystyrene, α -methyl-ethoxystyrene, α
 20 -methyl-chlorostyrene, α -methyl-bromostyrene;
 polyoxyethylene methacrylate,
 N,N,N-triethyl [2- (methacryloyloxy) ethyl] ammonium bromide,
 N,N,N-trimethyl [2- (methacryloyloxy) ethyl] ammonium
 chloride, N,N-diethyl-N-propyl [2- (methacryloyloxy)
 25 ethyl] ammonium bromide,
 N,N,N-trimethyl [3- (methacryloyloxy) -2-hydroxypropyl]
 ammonium chloride (QM) and N,N,N-trimethylammonium-
 methylstyrenebromide. Among them, methacrylic acid, stearyl
 methacrylate, benzyl methacrylate, buthyl methacrylate,

polyoxyethylene methacrylate and
N,N,N-trimethyl[3-(methacryloyloxy)-2-hydroxypropyl]
ammonium chloride (QM) are preferable, and benzyl
methacrylate is more preferable.

5 In the copolymer, a ratio of the monomer unit derived
from a monomer represented by the general formula [2], is
usually not less than 20 % and less than 100 %, preferably
30 to 95 %, more preferably 30-90 % and further more preferably
60 to 90 %.

10 As the polymer having a monomer unit derived from
a monomer represented by the general formula [2] in accordance
with the present invention, either a commercially available
polymer or a polymer synthesized according to the method
described in JP-A-10-45794 and JP-A-2000-239696 may be used.

15 The agglutination accelerator of the present
invention comprises a polymer having a monomer unit derived
from the monomer represented by the general formula [2],
as described hereinabove. The agglutination accelerator is
used, for example, by properly dissolving in various reagents
20 (usually in a form of solution) used in the known immunoassay,
in which a target substance is measured based on agglutination
and the like derived from an antigen-antibody reaction such
as immunoturbidimetry, immunonephelometry, latex
agglutination, preferably by dissolving in reagents used
25 in the immunoturbidimetry or the latex agglutination, and
more preferably by dissolving in reagents used in the latex
agglutination. Concentration of the agglutination
accelerator used in the various reagents is generally 0.1
to 20% by W/V, preferably 0.1 to 10% by W/V, more preferably

0.1 to 5% by W/V and further more preferably 0.1 to 2.0% by W/V, as a concentration in the reaction solution of the antigen-antibody reaction. The reaction under such concentration results in suppressing the increase of the blank value within a range not affecting the assay, and as a result, the agglutination accelerating effect of the present invention can be exhibited sufficiently.

In addition, the assay of the present invention may be performed by using various reagents used in the known immunoassay, in which a target substance is measured based on agglutination and the like derived from the antigen-antibody reaction such as immunoturbidimetry, immunonephelometry, latex agglutination, according to the known operation method, except that the assay is performed under the coexistence of the polymer of the present invention or the reagent kit of the present invention admixed at the above concentration on the antigen-antibody reaction.

Examples of buffers used in the assay of the present invention include every buffer generally used in the immuniturbidimetry and immunonephelometry such as Tris buffer, phosphate buffer, veronal buffer, borate buffer and Good's buffer. A pH in the assay is not particularly limited so long as it is within a range not suppressing the antigen-antibody reaction, and generally is suitably selected from a range of 6 to 10. A method measuring scattered light (nephelometry) can be performed, for example, according to the description in "Principles of Clinical Laboratory Tests" 30th Ed., 2nd Copy, p. 851-853 (1993), Kanehara Publ. Co. Ltd. A method measuring transmitted light

(immunoturbidimetry) can be performed, for example, according to the description in "Principles of Clinical Laboratory Tests" 30th Ed., 2nd Copy, p. 853-854 (1993), Kanehara Publ. Co. Ltd. The latex agglutination test, in
5 which a target substance is assayed based on the result obtained by measuring an extent of agglutination of latex sensitized (immobilized) with an antibody or an antigen to the target substance based on a change of scattered light or transmitted light, can be performed, for example,
10 according to the description in "New Actual Cases of Immunoassay, and Applications to Developments of Diagnostic Reagents and Drugs", p. 103-187, Keiei Kyoiku Publ. Co. Ltd.

The measurement of scattered light or transmitted light in the nephelometry, immunoturbidimetry and latex
15 agglutination can be performed by using a multipurpose biochemical analyzer such as automatic analyzer and spectrophotometer and a special-purpose apparatus for nephelometry such as laser nephelometry. Details can be provided by a manual of each apparatus.

20 The reagent kit of the present invention comprises a combination of the reagent containing a copolymer obtained by polymerizing the monomer represented by the general formula [2] and a monomer selected from a group consisting of acrylic acid or acrylate ester, methacrylic acid or
25 methacrylate ester, acrylamide or N-substituted derivatives thereof, methacrylamide or N-substituted derivatives thereof, and styrene or derivatives thereof, and the reagent containing an antibody to a prostate-specific antigen or a prostate-specific antigen. Specific examples

of the monomer represented by the general formula [2] and a monomer selected from a group consisting of acrylic acid or acrylate ester, methacrylic acid or methacrylate ester, acrylamide or N-substituted derivatives thereof, methacrylamide or N-substituted derivatives thereof, and styrene or derivatives thereof are described as above. Further, guanidine or arginine, which has an action to suppress non-specific reaction of the antigen-antibody reaction, may be added to the reagent containing the copolymer obtained by polymerizing the monomer represented by the general formula [2] and a monomer selected from a group consisting of acrylic acid or acrylate ester, methacrylic acid or methacrylate ester, acrylamide or N-substituted derivatives thereof, methacrylamide or N-substituted derivatives thereof, and styrene or derivatives thereof. Amount thereof to be added is not particularly limited so long as the intended effect can be obtained, and is preferably 100 to 700 mM for guanidine and 100 to 400 mM for arginine.

The antibody to prostate-specific antigen or the prostate-specific antigen in the reagent containing the antibody to prostate-specific antigen or the prostate-specific antigen may be fixed on a carrier or the like, and a preferable carrier is, for example, latex and the like.

Further, the above reagent kit may optionally contain a buffer (for example, Tris buffer, phosphate buffer, veronal buffer, borate buffer and Good's buffer), a stabilizer agent (for example, albumin, globulin, water-soluble gelatin, surface active agent, sugars), an

antiseptic (for example, salicylic acid, benzoic acid, sodium azide), and a reagent which are used in this field and do not inhibit stability of coexisting reagent or not inhibit the antigen-antibody reaction. Concentration
5 thereof in use may be within a range of the concentration generally used in this field.

As obvious from the above, the present invention provides an immunoassay of PSA using an agglutination accelerator, which has an agglutination accelerating effect
10 equal to or stronger than that of the conventionally known agglutination accelerator; hardly generates non-specific turbidity; and hardly generates salting out even in a solution with a high salt concentration, and a reagent kit comprising combining a reagent containing said agglutination
15 accelerator and the reagent containing an antibody to PSA or the PSA. Since the agglutination accelerator can exhibit an agglutination accelerating effect equal to or stronger than the action of the conventional means even in a high salt concentration, and makes the non-specific turbidity
20 difficult to occur, a PSA assay with high precision can be achieved.

The present invention will be explained by the following Examples, but the present invention is not limited by these Examples.

25

EXAMPLES

Synthesis Example 1: Synthesis of a copolymer of 2-methacryloyloxyethylphosphorylcholine (MPC) and n-butyl methacrylate (BMA) (8:2)

A glass reaction tube for polymerization was charged with 4.7 g (16 mM) of MPC and 0.57 g (4 mM) of BMA, then further added with 0.03 g of 2,2'-azobisisobutyronitrile (AIBN) as a polymerization initiator and 20 ml of methanol as a polymerization solvent. The air in the reaction tube was sufficiently exchanged with argon, and the reaction tube was sealed. Then polymerization reaction was conducted at 60 °C for 24 hours. The obtained reaction mixture was ice-cooled, then added with 400 ml of diethyl ether drop-wisely to precipitate the polymer. Said precipitate was filtered, washed sufficiently with diethyl ether and dried under reduced pressure to obtain a polymer as a white powder. The polymer thus obtained was named polymer 1. A molecular weight of polymer 1 was 600,000.

15

Synthesis Example 2: Synthesis of a copolymer of MPC/BMA (5:5)

A similar synthesis as in the above-described Synthesis Example 1 was repeated except that 20 mM in total of MPC and BMA were used in a molar ratio of 5:5, to obtain polymer 2. A molecular weight of polymer 2 was 338,000.

20

Synthesis Example 3: Synthesis of a copolymer of MPC/BMA (3:7)

A similar synthesis as in the above-described Synthesis Example 1 was repeated except that 20 mM in total of MPC and BMA were used in a molar ratio of 3:7, to obtain polymer 3. A molecular weight of polymer 3 was 92,000.

25

Synthesis Example 4: Synthesis of a copolymer of MPC/stearyl methacrylate (9:1)

A similar synthesis as in the above-described Synthesis Example 1 was repeated except that stearyl methacrylate was used instead of BMA and 20 mM in total of MPC and stearyl methacrylate were used in a molar ratio of 9:1, to obtain polymer 4. A molecular weight of polymer 4 was 130,000.

10 Synthesis Example 5: Synthesis of a copolymer of MPC/benzyl methacrylate (8:2)

A similar synthesis as in the above-described Synthesis Example 1 was repeated except that benzyl methacrylate was used instead of BMA and 20 mM in total of MPC and benzyl methacrylate were used in a molar ratio of 8:2, to obtain polymer 5. A molecular weight of polymer 5 was 240,000.

Synthesis Example 6: Synthesis of a homopolymer of MPC

20 A glass reaction tube for polymerization was charged with 5.9 g (20 mM) of MPC, then further added with 0.03 g of 2,2'-azobisisobutyronitrile (AIBN) as a polymerization initiator and 20 ml of methanol as a polymerization solvent. The air in said reaction tube was sufficiently exchanged with argon, and the reaction tube was sealed. Then 25 polymerization reaction was conducted at 50 °C for 24 hours. The reaction mixture was ice-cooled, then added with 400 ml of diethyl ether drop-wisely to precipitate the polymer. The precipitate obtained was filtered, washed sufficiently

with diethyl ether and dried under a reduced pressure to obtain a polymer as a white powder. The polymer thus obtained was named polymer 6. A molecular weight of polymer 6 was 110,000.

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Synthesis Example 7: Synthesis of a copolymer of MPC/QM (9:1)

A similar synthesis as in the above-described Synthesis Example 1 was repeated except that QM was used instead of BMA and 20 mM in total of MPC and QM were used in a molar ratio of 9:1, to obtain polymer 7. A molecular weight of polymer 7 was 33,000.

Synthesis Example 8: Synthesis of a copolymer of MPC/methacrylic acid (3:7)

A similar synthesis as in the above-described Synthesis Example 1 was repeated except that methacrylic acid was used instead of BMA and 20 mM in total of MPC and methacrylic acid were used in a molar ratio of 3:7, to obtain polymer 8. A molecular weight of polymer 8 was 288,000.

20

Example 1: Assay of prostate-specific antigen (PSA) by LIA (Effect of polymer type on the agglutination accelerating action)

(1) Preparation of an anti-human PSA antibody sensitized latex test solution

25

A mixture of 0.5 ml of 50 mM borate buffer (pH 7.1) containing 0.8 mg of anti-human PSA mouse monoclonal antibody (made by Wako Pure Chemical Industries, Ltd.) and 0.5 ml of 50 mM borate buffer (pH 7.1) containing a polystyrene

latex (particle size 0.22 μ m, made by Sekisui Chem. Co. Ltd.) suspended in 2% by W/V, was reacted at 25 °C for 2 hours. Subsequently, the latex separated by centrifugation was washed with 50 mM borate buffer (pH 7.1), and said latex
5 was suspended in 50 mM borate buffer (pH 7.3) containing 0.5% by W/V of BSA so that the concentration of the latex became 0.1% by W/V, and the thus obtained mixture was used as an anti-human PSA antibody sensitized (immobilized) latex test solution.

10 (2) Sample

A PSA derived from human seminal fluid (made by Wako Pure Chemical Industries, Ltd.) was dissolved in 10 mM phosphate buffer (0.85% NaCl) containing 1.0% by W/V of BSA, and the PSA solution of a predetermined concentration was
15 used as a sample.

(3) Reagents

i) Test solution No. 1

A solution of 100 mM HEPES-NaOH buffer (pH 7.0) containing 1.5% of a polymer as an agglutination accelerator,
20 0.1% of BSA and 2% of NaCl, or a solution of 100 mM HEPES-NaOH buffer (pH 7.0) containing 0.1% of BSA and 2% of NaCl as a reagent without agglutination accelerator was designated as test solution No. 1.

ii) Test solution No. 2

25 The anti-human PSA antibody sensitized (immobilized) latex test solution prepared in (1) was used as a test solution No. 2.

(4) Assay method

Assay was performed under the following measuring

conditions by using a BM-8 automatic analyzer made by JEOL Ltd.

Sample: 5 μ l

Test solution No. 1: 90 μ l

5 Test solution No. 2: 30 μ l

Assay method: 2 point end method (34-65)

Main wavelength: 571 nm

(5) Results

10 The values of absorbance (turbidity) obtained are shown in Table 1. The value in the Table was calculated by subtracting the absorbance value for the reagent blank from the value obtained for the test solution, and then multiplying by 10,000.

15 Comparative Example 1

The reagent as same as in Example 1 was used and the measurement as same as in Example 1 was repeated, except that the polymers of the present invention were replaced by PEG 6000 with a concentration of 1.5%.

20 The results obtained are shown in Table 1 together with the results obtained in Example 1.

Table 1

PSA (ng/mL)	Without Aggluti nation accelerator	Example 1			Comp. Example 1
		Polymer 1	Polymer 5	Polymer 6	PEG 6000
2	37	71	90	60	75
10	160	512	605	296	292

50	817	5483	5824	2440	1914
----	-----	------	------	------	------

As obvious from the results in Table 1, the agglutination accelerating action was observed in the assay of PSA, by using any polymer of the present invention. In addition, as compared with the results of PEG 6000, it can be understood that every polymer of the present invention could exhibit a higher agglutination accelerating action than that of PEG 6000, and among them the polymer 5 exhibited the highest effect in these 3 polymers.

10

Example 2: Assay of PSA by LIA (Effect of polymer concentration on agglutination accelerating action)

(1) Sample

A PSA derived from human seminal fluid (made by Wako Pure Chemical Industries, Ltd.) was dissolved in 10 mM phosphate buffer (0.85% NaCl) containing 1.0% by W/V of BSA, and the PSA solution with a predetermined concentration was used as a sample.

(2) Reagents

20 i) Test solution No. 1

A solution of 100 mM HEPES-NaOH buffer (pH 7.0) containing a predetermined concentration of the polymer 5 as an agglutination accelerator, 0.1% of BSA and 2% of NaCl, or a solution of 100 mM HEPES-NaOH buffer (pH 7.0) containing 0.1% of BSA and 2% of NaCl as a reagent without agglutination accelerator was designated as test solution No. 1.

25 ii) Test solution No. 2

The anti-human PSA antibody sensitized

(immobilized) latex test solution obtained and prepared in Example 1 was used as test solution No. 2.

(3) Assay method

Assay was performed under the following measuring
5 conditions by using a BM-8 automatic analyzer made by JEOL Ltd.

Sample: 5 μ l

Test solution No. 1: 90 μ l

Test solution No. 2: 30 μ l

10 Assay method: 2 point end method (34-65)

Main wavelength: 571 nm

(4) Results

The values of absorbance (turbidity) obtained are shown in Table 2. The value in the Table was calculated by
15 subtracting the absorbance value for the reagent blank from the value obtained for the test solution, then multiplying by 10,000.

Table 2

PSA (ng/mL)	Without Agglutination Accelerator	Example 2		
		1.0% Addition	1.5% Addition	2.0% Addition
2	37	58	90	833
10	160	318	605	3869
50	817	3096	5824	7454

As obvious from the results in Table 2, it can be
20 understood that the agglutination accelerator action of the polymer 5 in the assay by LIA is increased depending on the increase in the concentration of polymer 5.

Referential Example 1: Assay of CRP by latex
25 immunoturbidimetric assay (LIA)

(1) Preparation of an anti-human CRP antibody sensitized latex test solution

A mixture of 2 ml of 50 mM borate buffer (pH 7.1) containing 1.1 mg of anti-human CRP goat polyclonal antibody (made by International Immunology Corp.) and 1 ml of 50 mM borate buffer (pH 7.1) containing a polystyrene latex (particle size 0.12 μ m, made by Sekisui Chem. Co. Ltd.) suspended in 1% by W/V, was reacted at 30 °C for 2 hours. Subsequently, the latex separated by centrifugation was washed with 50 mM borate buffer (pH 7.1), and said latex was suspended in 50 mM borate buffer (pH 7.3) containing 0.5% by W/V of bovine serum albumin (BSA) so that the concentration of the latex became 0.2% by W/V. The thus obtained suspension was used as an anti-human CRP antibody sensitized (immobilized) latex test solution.

(2) Sample

Physiological saline solution (0.85% NaCl, CRP concentration: 0 mg/[[dL]]mL) was used as a sample for reagent blank assay, and solutions of CRP calibrator set (CRP concentration: 0.03 mg/[[dL]]mL, made by Wako Pure Chemical Industries, Ltd.) diluted into 10 steps of concentrations with physiological saline solution (0.85% NaCl) were used as samples for CRP-specific absorbance measurement.

(3) Reagents

i) Test solution No. 1

Previously synthesized polymer 1, polymer 5 and polymer 6 were used as the agglutination accelerator and a solution of 100 mM HEPES-NaOH buffer (pH 7.0) containing the predetermined concentration of the predetermined polymer,

0.1% of BSA and 1% of NaCl, or a solution of 100 mM HEPES-NaOH buffer (pH 7.0) containing 0.1% of BSA and 1% of NaCl as a reagent without agglutination accelerator, was designated as test solution No. 1.

5 ii) Test solution No. 2

The anti-human CRP antibody sensitized (immobilized) latex test solution prepared in the above (1) was used as test solution No. 2.

(4) Assay method

10 Assay was performed under the following measuring conditions by using a BM-8 automatic analyzer made by JEOL Ltd.

Sample: 1.25 μ l

Test solution No. 1: 75 μ l

15 Test solution No. 2: 25 μ l

Assay method: 2 point end method (34-65)

Main wavelength: 571 nm

(5) Results

20 The values of absorbance (turbidity) obtained are shown in Table 3. The value in the Table was calculated by subtracting the absorbance value of the reagent blank from the value obtained for the test solution, then multiplying by 10,000.

25 Comparative Example 2

The reagent as same as in Example 1 was used and the measurement as same as in Example 1 was repeated, except that the polymers of the present invention were replaced by PEG 6000 with the predetermined concentration, which was

generally used as an agglutination accelerator.

The results obtained are shown in Table 3 together with the results in Referential Example 1.

Table 3

CRP (mg/mL)	No Addition	Referential Example 1						Comparative Example 2	
		Polymer 1		Polymer 5		Polymer 6		PEG 6000	
Addition Conc'n	0	0.5%	1.0%	0.5%	1.0%	0.5%	1.0%	0.5%	1.0%
0.003	302	436	626	427	566	381	487	371	468
0.006	750	1143	1815	1067	1544	941	1220	922	1163
0.009	1310	2075	3600	1950	3024	1720	2266	1598	2056
0.012	1961	3221	5595	2965	4662	2566	3447	2405	3029
0.015	2723	4577	7367	4152	6230	3564	4673	3333	4096
0.018	3527	5906	8656	5367	7725	4657	6012	4254	5194
0.021	4346	7022	9242	6512	8672	5685	7042	5154	6100
0.024	5212	8017	9464	7506	9104	6511	7754	6066	6974
0.027	5991	8630	9512	8225	9421	7387	8424	6827	7612
0.030	6533	9011	9458	8690	9480	7919	8752	7356	7996

Table 3

CRP (mg/dL)	No Addition	Referential Example 1						Comparative Example 2	
		Polymer 1		Polymer 5		Polymer 6		PEG 6000	
Addition Conc'n	0	0.5%	1.0%	0.5%	1.0%	0.5%	1.0%	0.5%	1.0%
0.3	302	436	626	427	566	381	487	371	468
0.6	750	1143	1815	1067	1544	941	1220	922	1163
0.9	1310	2075	3600	1950	3024	1720	2266	1598	2056
1.2	1961	3221	5595	2965	4662	2566	3447	2405	3029
1.5	2723	4577	7367	4152	6230	3564	4673	3333	4096
1.8	3527	5906	8656	5367	7725	4657	6012	4254	5194
2.1	4346	7022	9242	6512	8672	5685	7042	5154	6100
2.4	5212	8017	9464	7506	9104	6511	7754	6066	6974
2.7	5991	8630	9512	8225	9421	7387	8424	6827	7612
3.0	6533	9011	9458	8690	9480	7919	8752	7356	7996

5

As obvious from the results in Table 3, the agglutination accelerating action was observed with any polymer of the present invention. In addition, as compared with the results of PEG 6000 in Comparative Example 2, it can be understood that every polymer of the present invention

could exhibit a higher agglutination accelerating action than that of PEG 6000, in particular, polymer 1 and polymer 5 exhibit higher effects.

5 Referential Example 2: Assay of CRP in human serum by LIA
(1) Sample

Human sera of 12 cases were used as samples. And physiological saline solution (0.85% NaCl) and a CRP calibrator set (CRP concentration: 0.01, 0.03, 0.05,
10 | 0.2[[0]] and 0.3[[0]] mg/[[dL]]mL, made by Wako Pure Chemical Industries, Ltd.) were used as samples for preparation of a calibration curve.

(2) Reagents

i) Test solution No. 1

15 A solution of 100 mM HEPES-NaOH buffer (pH 7.0) containing 0.5% of the predetermined polymer as the agglutination accelerator, 0.1% of BSA and 1% of NaCl was designated as test solution No. 1.

ii) Test solution No. 2

20 The anti-human CRP antibody sensitized (immobilized) latex test solution prepared in Example 1 (1) was used as test solution No. 2.

(3) Assay method

25 Absorbance measurements for samples were performed under the following measuring conditions by using a BM-8 automatic analyzer made by JEOL Ltd.

A value of absorbance obtained by measuring absorbance of physiological saline solution was used as a reagent blank value.

A calibration curve was prepared from the values obtained by subtracting the reagent blank values from the absorbance values obtained by measuring each standard solution of the CRP calibrator set, and the CRP concentration of each standard solution. Thereafter, the value obtained by subtracting the reagent blank value from the absorbance value obtained by measuring the sample was applied to the calibration curve to obtain a CRP concentration in the human serum.

Sample: 1.25 μ l
Test solution No. 1: 75 μ l
Test solution No. 2: 25 μ l
Assay method: 2 point end method (34-65)
Main wavelength: 571 nm

(4) Results

CRP concentrations (~~mg/dL~~ ng/mL) obtained are shown in Table 4.

Comparative Example 3: Assay of CRP concentration in human serum by LT Auto Wako

Measurement by the same method as Referential Example 2 was repeated except that LT Auto Wako (made by Wako Pure Chemical Industries, Ltd.) was used as a reagent. The thus obtained CRP concentrations (~~mg/dL~~ ng/mL) are shown in Table 4 together with the results of Referential Example 2.

Table 4

Serum Specimen	Referential Example 2 (mg/dL ng/mL)	Comparative Example 3 (mg/dL ng/mL)
----------------	--	--

	Polymer 1	Polymer 5	Polymer 6	LT Auto Wako
1	0.01 0.1	0.02 0.2	0.02 0.2	0.02 0.2
2	0.03 0.3	0.04 0.4	0.05 0.5	0.04 0.4
3	0.04 0.4	0.04 0.4	0.06 0.6	0.05 0.5
4	0.10 1.0	0.11 1.1	0.11 1.1	0.11 1.1
5	0.16 1.6	0.17 1.7	0.18 1.8	0.17 1.7
6	0.19 1.9	0.20 2.0	0.21 2.1	0.21 2.1
7	0.31 3.1	0.32 3.2	0.32 3.2	0.31 3.1
8	0.96 9.6	0.97 9.7	0.96 9.6	0.96 9.6
9	1.06 10.6	1.06 10.6	1.06 10.6	1.05 10.5
10	1.29 12.9	1.30 13.0	1.27 12.7	1.27 12.7
11	1.77 17.7	1.78 17.8	1.76 17.6	1.78 17.8
12	1.86 18.6	1.89 18.9	1.86 18.6	1.88 18.8
Average	0.65 6.5	0.66 6.6	0.65 6.5	0.65 6.5

As obvious from the results in Table 4, the assay results obtained by the method of the present invention are equivalent to the measured values using the LT Auto Wako as the reagent for a conventional assay method, and it can be understood that the assay method of the present invention shows a high correlation to the conventional assay method, in other words, an target substance can be assayed with a high precision without generating non-specific reaction by using the polymer of the present invention.

Referential Example 3: Assay of CRP by turbidimetric immunoassay (TIA) (Effect of polymer type on agglutination accelerating action)

(1) Sample

Physiological saline solution (0.85% NaCl) was used as a sample for reagent blank assay, and a CRP calibrator set (CRP concentration: 0.01, 0.03, 0.05, 0.2[[0]] and 0.3[[0]] mg/[[dL]]mL, made by Wako Pure Chemical Industries, Ltd.) was used as samples for the CRP-specific absorbance

measurement.

(2) Reagents

1) Test solution No. 1

A solution of 50 mM HEPPSO-NaOH buffer (pH 8.2) containing 1% of predetermined polymer as an agglutination accelerator and 1% of NaCl, or a solution of 50 mM HEPPSO-NaOH buffer (pH 8.2) containing 1% of NaCl as a reagent without agglutination accelerator was designated as test solution No. 1.

10 ii) Test solution No. 2

CRP α Auto Wako antibody solution (made by Wako Pure Chemical Industries, Ltd.) was used as test solution No. 2.

(3) Assay method

15 Assay was performed under the following measuring conditions by using an automatic analyzer 7150 made by Hitachi, Ltd.

Sample: 10 μ l

Test solution No. 1: 250 μ l

20 Test solution No. 2: 50 μ l

Assay method: 2 point end method (24-50)

Main wavelength: 340 nm

Sub-wavelength: 700 nm

(4) Results

25 The values of absorbance (turbidity) obtained are shown in Table 5. The column indicating CRP concentration of 0.01 to 0.3 [[0]] (mg/[[dL]]mL) in Table 5 shows the values obtained by subtracting the absorbance values of the reagent blank from the absorbance values obtained for the CRP

calibrator set, then multiplying by 10,000. Further, the column indicating reagent blank shows the values obtained by measuring reagent blank under the conditions so that an absorbance of the purified water becomes 0, then multiplying

5 by 10,000.

Table 5

CRP (mg/ml)	Without agglutination promoter	Referential Example 3							
		Polymer 1	Polymer 2	Polymer 3	Polymer 4	Polymer 5	Polymer 6	Polymer 7	Polymer 8
Reagent Blank	198	177	188	187	173	189	188	201	211
0.01	5	52	10	6	16	21	13	25	59
0.03	26	142	57	32	87	112	93	139	218
0.05	57	264	106	75	150	208	162	264	396
0.20	385	1476	723	456	976	1327	1174	1478	2110
0.30	668	2770	1261	876	1829	2388	2136	2545	3376

Table 5

CRP (mg/dl)	Without agglutination promoter	Referential Example 3							
		Polymer 1	Polymer 2	Polymer 3	Polymer 4	Polymer 5	Polymer 6	Polymer 7	Polymer 8
Reagent Blank	198	177	188	187	173	189	188	201	211
1	5	52	10	6	16	21	13	25	59
3	26	142	57	32	87	112	93	139	218
5	57	264	106	75	150	208	162	264	396
20	385	1476	723	456	976	1327	1174	1478	2110
30	668	2770	1261	876	1829	2388	2136	2545	3376

According to the results in Table 5, in an assay of CRP by TIA, the agglutination accelerating action was also observed with any polymer of the present invention. Further, in any polymer, the reagent blank values were equal to the cases using reagents without the agglutination accelerators, and non-specific reaction was not observed.

15 Referential Example 4: Assay of CRP by TIA (Effect of polymer concentration on agglutination accelerating action)

(1) Sample

Physiological saline solution (0.85% NaCl) was used as a sample for reagent blank assay, and a CRP calibrator set (CRP concentration: 0.01, 0.03, 0.05, 0.2[[0]] and
5 | 0.3[[0]] mg/[[dL]]mL, made by Wako Pure Chemical Industries, Ltd.) was used as samples for the CRP-specific absorbance measurement.

(2) Reagents

i) Test solution No. 1

10 | A solution of 50 mM HEPPSO-NaOH buffer (pH 8.2) containing the predetermined concentration of polymer 1 as an agglutination accelerator and 1% of NaCl was designated as test solution No. 1.

ii) Test solution No. 2

15 | CRP α Auto Wako antibody solution (made by Wako Pure Chemical Industries, Ltd.) was used as test solution No. 2.

(3) Assay method

20 | Assay was performed under the following measuring conditions by using an automatic analyzer 7150 made by Hitachi, Ltd.

Sample: 10 μ l

Test solution No. 1: 250 μ l

Test solution No. 2: 50 μ l

25 | Assay method: 2 point end method (24-50)

Main wavelength: 340 nm

Sub-wavelength: 700 nm

(4) Results

The values of absorbance (turbidity) obtained are

shown in Table 6. The column indicating CRP concentration of 0.01 to 0.03 [[30]] (mg/[[dL]]mL) in Table 5 shows the values obtained by subtracting the absorbance values of the reagent blank from the absorbance values obtained for the CRP calibrator set, then multiplying by 10,000. Further, the column indicating reagent blank shows the values obtained by measuring reagent blank under the conditions so that an absorbance of the purified water becomes 0, then multiplying by 10,000.

10

Table 6

CRP (mg/[[dL]]ml)	Referential Example 4			
	0%	0.5%	1%	2%
Reagent Blank	192	189	177	186
0.01	3	10	52	77
0.03	32	61	142	212
0.05	66	122	264	380
0.2[[0]]	403	842	1476	1845
0.3 [[30]]	717	1597	2770	3203

According to the results in Table 6, it can be understood that the agglutination accelerator action of polymer 1 in the assay by TIA is increased depending on the increase in the concentration of polymer 1. Further, the reagent blank values were almost constant even with increased polymer concentration, and non-specific reaction was not observed.

20 Referential Example 5: Assay of CRP in human serum

(1) Sample

Human sera of 26 cases were used as samples. Physiological saline solution (0.85% NaCl) and a CRP

calibrator set (CRP concentration: 0.01, 0.03, 0.05,
0.2[[[0]] and 0.3[[0]] mg/[[dL]]mL, made by Wako Pure
Chemical Industries, Ltd.) were used as samples for
preparation of a calibration curve.

5 (2) Reagents

i) Test solution No. 1

A solution of 50 mM HEPPSO-NaOH buffer (pH 8.2)
containing 2.5% of polymer 5 as an agglutination accelerator
and 1% of NaCl was designated as test solution No. 1.

10 ii) Test solution No. 2

CRP_Q Auto Wako antibody solution (made by Wako Pure
Chemical Industries, Ltd.) was used as test solution No.
2.

(3) Assay method

15 Assay was performed under the following measuring
conditions by using an automatic analyzer 7150, made by
Hitachi, Ltd. according to the assay method for CRP_Q Auto
Wako.

A value of absorbance obtained by measuring
20 absorbance of physiological saline solution was used as a
reagent blank value. A calibration curve was prepared from
the CRP concentration of each standard solution and the values
obtained by subtracting the reagent blank values from the
absorbance values obtained by measuring each standard
25 solution of the CRP calibrator set. Thereafter, the value
obtained by subtracting the reagent blank value from the
absorbance value obtained by measuring the sample was applied
to the calibration curve to obtain a CRP concentration in
the human serum.

Sample: 10 μ l

Test solution No. 1: 250 μ l

Test solution No. 2: 50 μ l

Assay method: 2 point end method (24-50)

5 Main wavelength: 340 nm

Sub-wavelength: 700 nm

Comparative Example 4: Assay of CRP concentration in human serum by CRPQ Auto Wako

10 Measurements by the same method as Referential Example 5 were repeated except that test solution No. 1 of CRPQ Auto Wako (made by Wako Pure Chemical Industries, Ltd.) was used as the test solution No. 1 of reagent.

15 A correlation between the CRP concentrations (mg/[dL])mL obtained and the CRP concentrations (mg/[dL])mL obtained in the above Referential Example 5 is shown in Fig. 1. In Fig. 1, the correlation equation was: $Y = 1.010X - 0.04$, and the correlation coefficient was: $r = 0.9999$.

20 As obvious from the above results, it can be understood that the assay results of CRP obtained by the method of the present invention show a high correlation to the assay results of CRP using the conventional assay method, CRPQ Auto Wako, in other words, a target substance can be
25 assayed with a high precision without generating non-specific reaction by using the polymer of the present invention.

Referential Example 6: Assay of rheumatoid factor (RF) by

TIA (Effect of polymer type on agglutination accelerating action)

(1) Sample

Physiological saline solution (0.85% NaCl) was used
5 as a sample for reagent blank assay, and a RF TIA calibrator set (RF concentration: 37, 71, 156, 310 and 498 IU/ml, made by Wako Pure Chemical Industries, Ltd.) was used as samples for the RF-specific absorbance measurements.

(2) Reagents

10 i) Test solution No. 1

A solution of 50 mM HEPES-NaOH buffer (pH 7.4) containing 1% of the predetermined polymer as an agglutination accelerator and 4% of NaCl, or a solution of 50 mM HEPES-NaOH buffer (pH 7.4) containing 4% of NaCl as
15 a reagent without agglutination accelerator was designated as test solution No. 1.

ii) Test solution No. 2

A RF-HA Test Wako RF reaction test solution (made by Wako Pure Chemical Industries, Ltd.) was designated as
20 test solution No. 2.

(3) Assay method

Assay was performed under the following measuring conditions by using an automatic analyzer 7150 made by Hitachi, Ltd.

25 Sample: 15 μ l

Test solution No. 1: 250 μ l

Test solution No. 2: 75 μ l

Assay method: 2 point end method (24-50)

Main wavelength: 340 nm

Sub-wavelength: 700 nm

(4) Results

The value of absorbance (turbidity) obtained are shown in Table 7. The columns indicating RF concentration 37 to 498 (IU/mL) in Table 7 shows the values obtained by subtracting the absorbance values of the reagent blank from the absorbance values obtained for the CRP calibrator set, then multiplying by 10,000. Further, the column indicating reagent blank shows the values obtained by measuring reagent blank under the conditions so that an absorbance of the purified water becomes 0, then multiplying by 10,000.

Table 7

RF (IU/mL)	Without Agglutination Promoter	Referential Example 6					
		Polymer 1	Polymer 2	Polymer 3	Polymer 4	Polymer 5	Polymer 6
Reagent Blank	111	128	133	138	132	121	114
37	3	8	4	4	10	15	2
71	9	89	15	20	25	71	40
156	133	464	215	183	316	458	392
310	794	1192	938	897	1043	1217	1166
498	1525	1902	1679	1643	1866	1928	1941

According to the results in Table 7, the agglutination accelerating action was observed in an assay of RF by TIA, by using any polymer of the present invention. Further, in every polymer, the reagent blank values were equal to the cases using reagents without the agglutination accelerator, and non-specific reaction was not observed.

Referential Example 7: Assay of rheumatoid factor (RF) by TIA (Polymer concentration and agglutination accelerating

action)

The same reagent as Referential Example 6 was used and the same measurements as Referential Example 6 were repeated except that the predetermined concentration of polymer 6 was used as an agglutination accelerator.

(1) Results

The values of absorbance (turbidity) obtained are shown in Table 8. The columns indicating RF concentration 37 to 498 (IU/mL) in Table 8 show the values obtained by subtracting the absorbance values of the reagent blank from the absorbance values obtained for the CRP calibrator set, then multiplying by 10,000. Further, the column indicating reagent blank shows the values obtained by measuring reagent blank under the conditions so that an absorbance of the purified water becomes 0, then multiplying by 10,000.

Table 8

RF (IU/ml)	Referential Example 6				
	0%	0.5%	1%	2%	3%
Reagent Blank	111	118	114	110	97
37	3	3	2	32	156
71	9	15	40	189	377
156	133	254	392	654	872
310	794	1042	1166	1466	1707
498	1525	1997	1941	2223	2495

According to the results in Table 8, it can be understood that the agglutination accelerator action of polymer 6 in the assay by TIA is increased depending on the increase in the concentration of polymer 6. Further, the reagent blank values were almost constant even with increased polymer concentration, and non-specific reaction was not

observed.